Withanolides Q and R, Two New 23-Hydroxy-steroidal Lactones

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Two new steroidal lactones of the withanolide type, isolated from *Withania somnifera* Dun (Solanaceae), are characterized by the unusual presence of a C-23 hydroxy-group. Withanolide Q (1a) is $(22S,23R)-17\alpha,23,27$ -trihydroxy-1-oxowitha-2,5,24-trienolide and withanolide R (2a) is $(22S,23R)-6\alpha,7\alpha$ -epoxy-5 α ,23-dihydroxy-1-oxowitha-2,24-dienolide.

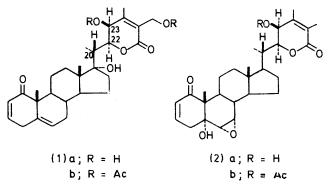
IN a previous paper ¹ we dealt with the structures of eleven steroidal lactones of the withanolide series, isolated from the 'Indian I' chemotype of *Withania somnifera* Dun. The twelfth compound of this group, $C_{28}H_{36}O_6$, isolated in minute quantities, was not at that time identified completely.

We have investigated recently the steroidal lactones produced in the offspring obtained by cross-pollination of the various chemotypes of *Withania somnifera*.² Crossing chemotypes I and III (both of which grow naturally in Israel) resulted in offspring F_1 , F_2 , and F_3 , each containing an array of withanolides. Chromatography of the crude extract obtained from the leaves of one of the offspring F_3 afforded, *inter alia*, a new withanolide which proved to be structurally related to the twelfth compound of the Indian chemotype. The present work is concerned with the structures of these two compounds, designated withanolides Q and R.

Withanolide Q (1a) exhibits two i.r. bands in the carbonyl region at 1 650 ($\alpha\beta$ -unsaturated ketone) and 1 705 cm⁻¹ ($\alpha\beta$ -unsaturated δ -lactone) and a u.v. maximum absorption at 218 nm (ϵ 17 900) attributable to the same unsaturated carbonyl chromophores. N.m.r. signals in the low-field region for three vinylic protons agree exactly with those displayed by withanolide G,³ thus allowing the assignment of a 2,5-dien-1-one structure to the AB ring system. An unresolved band at δ 4.42 accounting for four protons and a singlet at δ 2.11 for a vinylic methyl group are assigned to the side chain. ¹ I. Kirson, E. Glotter, D. Lavie, and A. Abraham, J. Chem.

Soc. (C), 1971, 2032.
² D. Lavie, Nobel Symposium 25, Chemistry in Botanical Classification, Academic Press, New York and London, 1973, p. 181.

Acetylation of (1a) gave the diacetate (1b), in the n.m.r. spectrum of which the δ 4.42 band was completely resolved into a two-proton singlet at δ 4.95 (27-H₂), a one-proton double doublet at δ 4.80 (22-H), and a one-proton doublet at δ 5.8 (23-H). The δ 4.95 singlet, in



conjunction with the single vinylic methyl signal (& 2.11) defines the substituents at C-24 and C-25 as CH₃ and CH₂·OAc. The double doublet at & 4.80 points to two protons only in the neighbourhood of the 22-H; however, in contrast to the 22-H in the 20-hydroxywithanolides ^{1,3} the signal of which is split by the two C-23 protons with J 12 and 5 Hz, in the present case the coupling constants are only 3.5 and 2 Hz, excluding any axial-axial interaction. Deuteriation experiments on deoxydihydrowithaferin A⁴ led to the exchange of the two C-23 protons and resulted in the appearance of the 22-H signal as a doublet (J 4 Hz). The only possible con-⁸ E. Glotter, I. Kirson, A. Abraham, and D. Lavie, *Tetrahedron*, 1973, 29, 1353.

⁴ D. Lavie, E. Glotter, and Y. Shvo, J. Chem. Soc., 1965, 7517.

clusion is that in (1b) the axial proton at C-23 is replaced by an acetoxy-group, thus explaining also the doublet at δ 5.81 (I 2 Hz) for the 23-H.

Double-resonance measurements confirmed the above interpretation: irradiation at δ 2.34 (20-H) leads to decoupling of the 21-H (singlet instead of doublet) and of the 22-H (doublet, J 2 Hz, instead of double doublet); conversely irradiation at δ 4.80 (22-H) leads to decoupling of the 23-H (singlet instead of doublet) and of the

protons (21-H₃ and 22-H) which are near the 17-OH are shifted downfield. The Cotton effect in the c.d. spectrum of the lactone ring in (1a) is positive, as in all other withanolides (260.5 nm; $\Delta \varepsilon + 3.85$). Consequently (1a) is (22S,23R)-17 α ,23,27-trihydroxy-1-oxowitha-2,5,-24-trienolide.

On the basis of the above data we can now assign a similar structure to the side chain of (2b) [the naturally occurring alcohol (2a) could not be isolated as such, but

N.m.r. signals * of withanolides Q and R and their acetates

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Compd.	2-H	3-H	6-H	7-H	22-H	23-H	$19-H_3$	$18-H_3$	$21-H_8$	Other
(la)	5.92dq	6.86dq	5.63		4.42	4.42	1.25s	0.87s	1.13d	22-H, 23-H, 27-H ₂ : broad
• •	$(10, 3, \bar{1})$	(10, 5, 2.5)	W1 8						(7)	band at 4.42; 28-H:
	(-, -, ,	(, , , , , ,	2		[4.83]		[1.22]	[0.78]	[1.26]	2.11s [2.31]
(1b)	5.90da	6.85da	5.63		4.80dd	5.81d	1.24s	0.79s	0.95d	27-H ₂ : 4.95s; 28-H ₃ and
()	(10, 3, 1)	(10, 5, 2.5)	W1 8		(2.5, 2)	(2)			(7)	acetates: 2.06, 2.09,
	(, -, -,	(,	2		[5.21]	、 /	[1.21]	[0.78]	[1.21]	2.16
(2b)	5.85da	6.56da	3.05d	3.33dd	4.73dd	5.63d	1.17s	0.80s	0.93d	27- and 28-H ₃ : 1.97; AcO:
()	(10,3,1)	(10, 4.5, 3)	(4)	(4, 1)	(3.5, 2)	(2)			(7)	2.15
	(=-,-,-)	· · · · · · · · · · · · · · · · · · ·	· ·	· · -/		• •			• •	

* δ Values: J/Hz in parentheses; data for solutions in C_5D_5N in square brackets.

21-Me; finally, irradiation at δ 5.81 (23-H) induces the decoupling of the 22-H (now doublet, J 3.5 Hz).

Treatment of (1b) with basic alumina at room temperature caused elimination of the 23-acetoxy-group, to give the corresponding pyrone, λ_{max} . 307 nm.

On the basis of the molecular weight of (1a), the compound must possess a third hydroxy-group which, in the absence of any relevant n.m.r. signal should be tertiary. It is assigned the 17α -position by analogy with other withanolides and in agreement with pyridine-induced n.m.r. shifts $\Delta(\text{CDCl}_3 - \text{C}_5\text{D}_5\text{N})$ of $18\text{-H}_3 + 0.01$, $21\text{-H}_3 - 0.26$, and 22-H - 0.41 p.p.m.¹

Comparison of the n.m.r. spectra of (1a) and its diacetate (Ib) (in CDCl₂) reveals two relevant aspects concerning the protons near to the 23-substituent. In (1b), the 18-H₃ and the 21-H₃ signals are shifted upfield, whereas the 22-H signal is shifted downfield by 0.40 p.p.m. A model shows that the 23-acetate group restricts rotation about the C(20)-C(22) bond; as a result, in the most preferred conformation, the 18- and the 21-protons are more distant from the 23-substituent than in (1a), whereas the 22-H is held closer to the 17α -OH, thus inducing deshielding of the latter and shielding of the former. The spectra of (1a) and (1b) in C_5D_5N support such an interpretation. Whereas in (1a) $\Delta(\text{CDCl}_3 - \text{C}_5\text{D}_5\text{N})$ is positive for 18-H₃ and negative for 21-H₃, 22-H, and 28-H₃, in (1b) 18-H₃ and 28-H₃ are practically unaffected, whereas the 21-H₃ and the 22-H signals are shifted downfield. It is known⁵ that the effect of pyridine is due to hydrogen bonding with the hydroxylic oxygens. In (1a) the pyridine coordinated to the 23-OH restricts rotation about the C(20)-C(22) bond in such a way that the distance between the 23-OH and the $18-H_3$ is increased (upfield shift of the $18-H_3$ signal); concomitantly, the protons which are close to the 23-OH and the 17-OH are significantly deshielded. In (1b), where pyridine does not co-ordinate around the 23-acetoxy-group, only the signals of those

⁵ P. V. Demarco, E. Farkas, D. Doddrell, B. L. Mylari, and E. Wenkert, J. Amer. Chem. Soc., 1968, **90**, 5840. only as the 23-monoacetate]. Comparison of n.m.r. signals confirms that the only difference between the side chains in (1b) and (2b) is the absence of the 27-OH in the latter. As to the substitution pattern of rings A and B, and the similarity to $(22R)-6\alpha,7\alpha$ -epoxy- $5\alpha,17\alpha$ -dihydroxy-1-oxowitha-2,24-dienolide¹ leaves no doubt, about the structure (2b). Since the molecular weight of (2b) indicates the presence of only seven oxygen atoms, the compound possesses only one tertiary OH (5α -OH).

The mass spectral fragmentation pattern of (1a) $(M^+ 470)$ supports the assigned structure. The base peak $(m/e\ 267)$ is due to the cleavage of the C(17)-C(20) bond, accompanied by elimination of a molecule of water. Cleavage of the C(20)-C(22) bond, which is the dominant process in the fragmentation of withanolides unsubstituted at C-23, is less important in the present case and leads to the signal $m/e\ 295\ (M^+ - 157\ - 18)$; the second possible fragment from this rupture, which would correspond to the lactone ring, is of low abundance. Nevertheless, there are two significant signals $(m/e\ 342$ and 324) related to the cleavage of the lactone itself along the C(22)-C(23) and the O-C:O bonds.

In the spectrum of the diacetate (1b) $(M^+ 554)$ the main signals correspond to the trivial elimination of water and acetic acid (m/e 536, 494, 476, 434, and 416). Loss of the whole side chain (accompanied by elimination of water) leads to the same fragment, m/e 267, as in (1a).

EXPERIMENTAL

M.p.s. were taken on a Fisher-Johns apparatus. Optical rotations were recorded with an automatic Perkin-Elmer 141 polarimeter and refer to solutions in chloroform. I.r. spectra were recorded on a Perkin-Elmer Infracord 137 spectrophotometer equipped with a sodium chloride prism, and refer to KBr pellets; u.v. spectra were recorded on a Cary 14 instrument (ethanol as solvent); n.m.r. spectra were determined on a Varian A-60 spectrometer for 5-10% solutions in deuteriochloroform, containing tetramethylsilane as internal standard. T.l.c. was carried out on chromatoplates of silica gel G (Merck) and spots were developed with iodine vapour. Mass spectra were taken

under the direction of Dr. Z. Zaretskii with an Atlas CH4 instrument. Analyses were performed in the microanalytical laboratory of the Weizmann Institute, under the direction of Mr. R. Heller.

Plant Material.—Reciprocal cross-pollination of Withania somnifera Dun., chemotypes I and III, resulted in the offspring F_1 . Self-pollination of F_1 afforded several offspring F_2 in which recombination of the chromosomes had taken place. One of these, which proved to be a new, artificial chemotype, remained unchanged when it was selfpollinated again. The offspring F_3 thus obtained constituted the plant material of the present work.

Isolation.—Crushed air-dried leaves (2 kg) were exhaustively extracted with methanol and the extract was worked up as previously described.³ The crude product (27 g) was chromatographed on a silica gel column (Woelm 0.063—0.1 mm; 3 kg). Elution with benzene–ethyl acetate (4.5:5.5) afforded crude withanolide Q (400 mg), which was purified on preparative chromatoplates (Woelm 60 F₂₅₄; 2 mm thick). Withanolide Q (1a) (120 mg) (22S,23R)-17 α ,23,27trihydroxy-1-oxowitha-2,5,24-trienolide] had m.p. 200—202° (from ethyl acetate), $[\alpha]_D - 6.6^\circ$ (c 0.12); ν_{max} 1 650 and 1 705 cm⁻¹, λ_{max} 218 nm (ϵ 17 950); c.d. (ethanol; c 0.63) 340 ($\Delta \epsilon$ -3.14) and 260.5 nm (+3.85) (Found: C, 71.45; H, 8.2%; M⁺, 470. C₂₈H₃₈O₆ requires C, 71.45; H, 8.15%; M, 470). Acetylation of (1a) (50 mg) with acetic anhydride in pyridine, overnight at room temperature, gave the 23,27diacetate (1b), m.p. 160-162° (from acetone), $[\alpha]_D = 88.8°$ (c 0.08); ν_{max} 1655, 1715, and 1745 cm⁻¹, λ_{max} 220 nm (ϵ 18 500) (Found: C, 69.3; H, 7.7%; M^+ , 554. C₃₂H₄₂O₈ requires C, 69.3; H, 7.65%; M, 554).

Withanolide R Monoacetate (2b).—Chromatography of the crude extract from leaves of W. somnifera 'Indian I' chemotype ¹ resulted in the separation of several pure withanolides and in two inseparable mixtures designated A and B. Mixture A was acetylated with acetic anhydride-pyridine overnight at room temperature and the crude mixture of acetates (0.5 g) was separated by thick-layer chromatography on silica gel PF₂₅₄ (1 mm thick in benzene-ethyl acetate (2:8). Extraction of the third band from the top afforded (22S,23R)-23-acetoxy-6 α ,7 α -epoxy-5 α -hydroxy-1-oxowitha-2,24-dienolide (2b) (22 mg), m.p. 182—184° (from acetone-hexane), [α]_D — 64° (c 0.09), ν_{max} . 1 698, 1 712, and 1 745 cm⁻¹, λ_{max} . 221.5 nm (ε 17 400) (Found: C, 70.6; H, 7.4%; M⁺, 510. C₃₀H₃₈O₇ requires C, 70.55; H, 7.5%; M, 510).

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